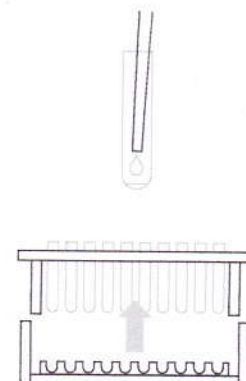
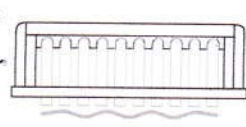

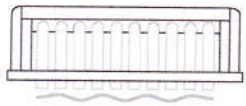

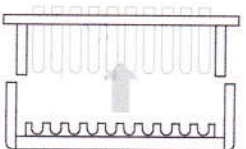


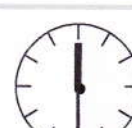

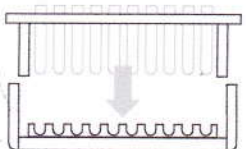


Mag particle

GLYPHOSATE DETAILED FLOWCHART

<p>1.</p> 	<p>Remove upper rack from magnetic base. Label test tubes for Standards, Control, and Samples.</p> <table border="1"> <thead> <tr> <th>Tube #</th> <th>Content</th> </tr> </thead> <tbody> <tr> <td>1, 2</td> <td>Diluent/Zero</td> </tr> <tr> <td>3, 4</td> <td>Standard 0 ppb</td> </tr> <tr> <td>5, 6</td> <td>Standard 1, 0.15 ppb</td> </tr> <tr> <td>7, 8</td> <td>Standard 2, 1.0 ppb</td> </tr> <tr> <td>9, 10</td> <td>Standard 3, 5.0 ppb</td> </tr> <tr> <td>11, 12</td> <td>Control</td> </tr> <tr> <td>13, 14</td> <td>Sample 1</td> </tr> <tr> <td>15, 16</td> <td>Sample 2</td> </tr> </tbody> </table> <p>there are 5 standards in kit</p> <p>0.075 0.2 0.75 4.0</p> <p>Add 300 μL of either Derivatized Standards, Control or Samples to the bottom of each test tube by inserting the pipette tip all the way into the bottom of the tube without touching the sides of the tube.</p> <p>want entire sample in bottom, none on tube walls.</p>	Tube #	Content	1, 2	Diluent/Zero	3, 4	Standard 0 ppb	5, 6	Standard 1, 0.15 ppb	7, 8	Standard 2, 1.0 ppb	9, 10	Standard 3, 5.0 ppb	11, 12	Control	13, 14	Sample 1	15, 16	Sample 2	<p>7.</p>  <p>Do not separate upper rack from lower base. Using a smooth motion, invert the combined rack assembly over a sink and pour out the tube contents; keep inverted and gently blot the test tube rims on several layers of paper toweling.</p>
Tube #	Content																			
1, 2	Diluent/Zero																			
3, 4	Standard 0 ppb																			
5, 6	Standard 1, 0.15 ppb																			
7, 8	Standard 2, 1.0 ppb																			
9, 10	Standard 3, 5.0 ppb																			
11, 12	Control																			
13, 14	Sample 1																			
15, 16	Sample 2																			
<p>2.</p>  <p>25 mL tip</p>	<p>Add 500 μL of thoroughly mixed Glyphosate Antibody Coupled Magnetic Particles down the inside wall of each tube by using the technique described in Box #1. <u>Vortex</u> for 1 to 2 seconds (at <u>low speed</u> to minimize foaming).</p>	<p>8.</p>  <p>Add 1 mL of Washing Solution down the inside wall of each tube by using the technique described in Box #7. Wait 2 minutes. Using a smooth motion, invert the combined rack assembly over a sink and pour out the tube contents; keep inverted and gently blot the test tube rims on several layers of paper toweling. Repeat this step two times.</p> <p>500 or 25 mL tip</p>																		
<p>3.</p> 	<p>React 30 minutes at room temperature (15° - 30°C).</p>	<p>9.</p>  <p>Lift the upper rack (with its tubes) off the magnetic base; add 500 μL of Color Reagent down the inside wall of each tube by using the technique described in Box #4. <u>Vortex</u> for 1 to 2 seconds (at low speed to minimize foaming).</p> <p>5 mL, 10 mL, 25 mL tip</p>																		
<p>4.</p>  <p>25 mL tip</p>	<p>Add 250 μL of Glyphosate Enzyme Conjugate down the inside wall of each tube by aiming the pipet tip 1/4" to 1/2" below the tube rim without touching the rim or tube wall with the pipet tip; deliver liquid gently.</p> <p>Vortex Mix: 1-2 sec.</p>	<p>10.</p>  <p>React for 20 minutes at room temperature (15° - 30° C). During this period, add 1 mL of Washing Solution into a clean tube for use as an instrument blank in Step 10.</p>																		
<p>5.</p> 	<p>React 30 minutes at room temperature (15° - 30°C).</p>	<p>11.</p>  <p>Add 500 μL of Stopping Solution down the inside wall of each tube by using the technique previously described. Read results at 450 nm within 15 minutes after adding the Stopping Solution. Multiply results of samples by the appropriate dilution factor (if any).</p> <p>n.o. mixing</p> <p>5 mL, 10 mL, 25 mL tip</p>																		
<p>6.</p> 	<p>Combine the upper rack with the magnetic base; press all tubes into base; allow 2 minutes for the particles to separate.</p>	<p>[Safety Caution: Stopping Solution contains diluted sulfuric acid.]</p>																		

For Ordering or Technical Assistance Contact:
 ABRAXIS, LLC 54 Steamwhistle Drive, Warminster, PA 18974
 Phone: 215-357-3911 Fax: 215-357-5232
 Web: www.abraxiskits.com

Glyphosate Magnetic Particle Kit Part # 500080, 120 Test

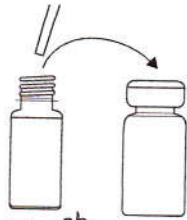


Glyphosate ELISA, Derivatization Procedure

this uses a different set of test tubes than assaying.

1. Derivatization Reagent Preparation

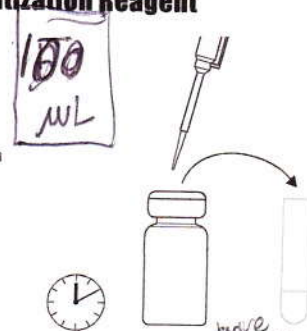
Dilute the derivatization reagent by adding 3.5 mL of the Derivatization Reagent Diluent (clear screw top glass vial) to the derivatization reagent vial (clear crimp top glass vial). Vortex and set aside.



Vortex 10 times ^{briefly} at high speed.

4. Addition of Derivatization Reagent

Add 100 uL of the diluted derivatization reagent (prepared in step 1) to each standard, control, and sample successively using a micropipette. Vortex each tube immediately after the addition of derivatization reagent for 15-30 seconds. Incubate tubes at room temperature for 10 minutes.



5 mL tip partly full

Vortexing: seven times or until swirls are gone. High speed.

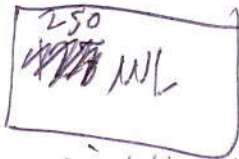
2. Addition of Sample to Test Tubes

Add 250 uL of each standard, control, and sample to the appropriate labeled ~~glass~~ test tube.



1.35 mL total volume

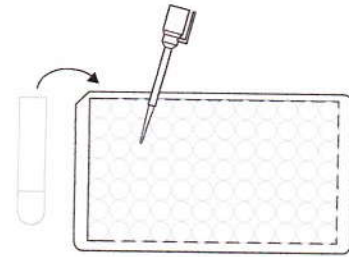
plastic



adjustable single shot pipettor

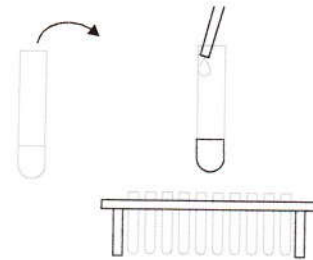
5. Analysis by ELISA

The derivatized standards, control, and samples can then be analyzed using the Glyphosate Plate or Tube ELISA Kits.



[see separate flow chart]

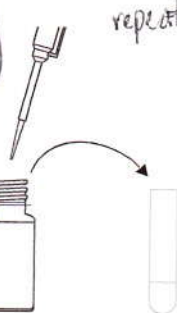
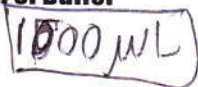
OR



3. Addition of Buffer

Add 1 mL of Glyphosate assay buffer to each tube. Vortex each tube for approximately 1-2 seconds.

50 or 25 mL repeater tip



Vortex each tube ~~just~~ just after adding the assay buffer.

High speed, touch four times.

For Ordering or Technical Assistance Contact:
 ABRAXIS, LLC
 54 Steamwhistle Drive
 Warminster, PA 18974
 Phone: 215-357-3911
 Fax: 215-357-5232
 www.abraxiskits.com



Glyphosate Derivatization Kits Part#'s 500084 & 500087

use the 25 mL pipettor tip or 50 mL if > 23 tubes

one tube per sample or standard, except for "C" tubes